

## NEURAL NETWORK APPROACH FOR ESTIMATION OF LACTATE

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**Abstract**

Lactate is cleared from the body by transporting it to the liver. This process of the body requires adequate amounts of oxygen. Due to certain ailments such as heart failure, respiratory dysfunctions, or severe infection, there is a shortage of oxygen supply. This leads to inefficient elimination of lactate and hence abnormal accumulation of lactate in the blood. To provide timely intervention, lactate needs to be monitored in a non-invasive manner in a critical care scenario. This manuscript describes a method of predicting lactate using fixed wavelength in the near-infrared region. We have selected wavelengths namely 2299, 2285, 2259, 2225, and 2129nm which correspond to the absorption peaks and valleys of lactate. We have performed a comparative analysis of partial least square regression and principal component analysis-artificial neural network for regression analysis. We obtained a root mean square error of 2.02 mg/dL with partial least square regression and a root mean square error of 0.15 mg/dL with principal component analysis-artificial neural network. Hence, the artificial neural network can be employed to predict lactate for medical applications.

There is a buildup of lactate in the body when it generates the required energy by the anaerobic pathway. The body clears the tissue lactate by carrying it to the liver where it either gets oxidized to carbon dioxide or transformed to glucose through cyclic process. This process requires adequate amounts of oxygen supply. However in conditions such as hypoxia where there is a shortage of oxygen due to underlying conditions such as heart failure, respiratory dysfunctions, or severe infection. In such conditions, lactate elimination is hindered. This leads to the accumulation of lactate in the blood beyond normal allowable levels. The normal levels in the human blood of lactate are less than 2mmol/L (18.02mg/dL). The above condition can lead to lactic acidosis due to altered pH levels as a result of high lactate concentrations in the blood. There can be other life-threatening conditions such as difficulty in breathing, confusion, and even coma [1]. Lactate concentrations higher than 4 mmol/L have been found in myocardial infarction [2], cardiac arrest [3], circulatory failure [4,5], and emergency trauma situations [6,7]. Hence a robust, continuous and non invasive method is of paramount importance in critical care scenarios.

Earlier investigations have shown the viability of Near Infrared (NIR) radiation for in-vitro lactate prediction [8,9]. In one study, ultraviolet (UV)/visible, NIR and Mid Infrared (MIR) radiation were used to estimate lactate contained in phosphate-buffered saline (PBS) in the concentration range of 0 to 20 mmol/L. Regression analysis were done using Partial Least Squares Regression (PLSR) and leave-one-out cross-validation gave a root mean square error of cross-validation of 1.59, 0.89, and 0.49 mmol/L for UV/visible, NIR, and MIR region respectively [10]. In this work we are attempting to bring out the efficacy near infrared region for lactate estimation by using reduced wavelengths instead of the entire absorption spectra. Earlier research works have reported absorption signature of lactate in combination band [11,12]. This work carefully selects wavelengths at the reported absorption peaks and valleys which lactate has. These wavelengths are found to correlate with concentration changes in lactate in the sample. The selected wavelengths are 2299, 2285, 2259, 2225, 2129nm. Reduction of wavelengths points reduces the computing and memory resources required to build estimation models.

Near infrared spectra were recorded of 64 laboratory samples in the range of 2050-2350nm using spectrophotometer (Model V-770 by Jasco, Japan). The laboratory samples contained glucose, ascorbate, urea, lactate, and alanine in aqueous solution. These above compounds were used as they resemble the blood tissue. The concentration of alanine was made to vary from 10 to 28 mg/dL, concentration of urea was made to vary from 11 to 20 mg/dL, concentration of lactate was made to vary from 12 to 22 mg/dL, concentration of glucose was made to vary from 70 to 280 mg/dL and concentration of ascorbate was made to vary from 2 to 5 mg/dL. All the compounds used were of analytical grade (Sigma Aldrich Ltd.). A quartz cuvette was used as a sample holder which had a path length of 1mm.

Partial least square regression (PLSR) and Principal Component Analysis Artificial Neural Network (PCA-ANN) were used for regression analysis. For the first regression analysis PLSR was implemented on the recorded data using ParLeS 3.1 software [13], which uses orthogonalized PLSR algorithm. The second regression analysis PCA-ANN, needs PCA algorithm to be applied on the input data which is done using MATLAB R2021b. The data is centered before passing through PCA algorithm. The Statistics and Machine Learning Toolbox necessary to run the PCA in MATLAB R2021b. The toolbox leverages the Singular value decomposition (SVD)

algorithm to implement the PCA. The preprocessed data by PCA is fed as input to the ANN for the purpose of machinelearning. The algorithm used to train the network is Levenberg-Marquardtback propagation algorithm. This is a part of the neural network fitting application which is bundled along with MATLAB R2021b. The tan-sigmoid transferfunction is used as an activation function for the hidden layer and the linearactivation function is used in the output layer. The created ANN is a shallownetwork, as it contains only one hidden layer, an input layer and an output layer.The scores on the principal components are given as input to the neural network. The input layer contains 3 neurons, hidden layer contains 5 neurons andthe output layer contains one neuron. For Both the Regression analysis, 57 ofthe 64 samples were used as calibration/training dataset. The rest 7 sampleswere used for validation of the created models where lactate was estimated inthe unknown samples.

The table 1 shows the analysis done using the above outlined methods. Thefirst approach using PLSR we extracted 4 factors to model the data. TheRoot Mean Square Error(RMSE) for prediction of lactate on the 7 validationsamples was 2.02 mg/dL. In the second approach, namely PCA-ANN 3 principalcomponents were extracted from the input spectral data which explained 99.97%of variance. These were then fed as input to the ANN. The lactate estimationon the validation set gave a RMSE of 0.15 mg/dL. Table 1: Estimation of lactate using the PLSR and PCA-ANN The PLSR method employed above is a multivariate method which models the linear relationships between the block of predictor variables (absorption signatures) andthe block of response variables (sample concentrations). However, there maybe many underlying non-linear effects occurring in the sample such as nonlinear detector response, stray light, sample turbidity, and multiple scattering due toin homogeneities in the sample, shifts in the width and positions the absorption bands due to variations in sample temperature and composition [14]. ThePCA-ANN is able to model the linear as well as the non-linear relationshipbetween predictor variables and response variables. The above results indicatethat PCA-ANN approach enhances the accuracy of estimation methodology andis a superior method as compared to PLSR.

| Sr. no. | Actual Concentration in mg/dL |         |           |         |         | Estimated lactate by PCA-ANN in mg/dL | Estimated lactate by PLSR in mg/dL |
|---------|-------------------------------|---------|-----------|---------|---------|---------------------------------------|------------------------------------|
|         | Urea                          | Glucose | Ascorbate | Aniline | Lactate |                                       |                                    |
| 1       | 20                            | 70      | 5         | 28      | 12      | 11.93                                 | 13.96                              |
| 2       | 20                            | 100     | 5         | 10      | 12      | 12.09                                 | 14.27                              |
| 3       | 11                            | 200     | 2         | 10      | 22      | 22.12                                 | 20.12                              |
| 4       | 11                            | 280     | 5         | 28      | 12      | 11.94                                 | 14.76                              |
| 5       | 20                            | 280     | 5         | 28      | 22      | 21.69                                 | 22.93                              |
| 6       | 20                            | 200     | 2         | 28      | 12      | 12.16                                 | 11.46                              |
| 7       | 11                            | 100     | 5         | 10      | 12      | 12.08                                 | 14.68                              |

## REFERENCES

- [1] M. Garcia-Alvarez, P. Marik, and R. Bellomo, "Sepsis-associated hyperlactatemia," *Crit Care*, vol. 18, no. 5, p. 503, Oct. 2014, doi: 10.1186/s13054-014-0503-3.
- [2] R. J. Henning, M. H. Weil, and F. Weiner, "Blood lactate as prognostic indicator of survival in patients with acute myocardial infarction," *Circ Shock*, vol. 9, no. 3, pp. 307–315, 1982.
- [3] M. H. Weil, C. E. Ruiz, S. Michaels, and E. C. Rackow, "Acid-base determinants of survival after cardiopulmonary resuscitation:," *Critical Care Medicine*, vol. 13, no. 11, pp. 888–892, Nov. 1985, doi: 10.1097/00003246-198511000-00005.
- [4] G. Broder and M. H. Weil, "Excess Lactate: An Index of Reversibility of Shock in Human Patients," *Science*, vol. 143, no. 3613, pp. 1457–1459, Mar. 1964, doi: 10.1126/science.143.3613.1457.
- [5] M. H. Weil and A. A. Aff, "Experimental and Clinical Studies on Lactate and Pyruvate as Indicators of the Severity of Acute Circulatory Failure (Shock)," *Circulation*, vol. 41, no. 6, pp. 989–1001, Jun. 1970, doi:10.1161/01.CIR.41.6.989.
- [6] J. Aduen, "The Use and Clinical Importance of a Substrate-Specific Electrode for Rapid Determination of Blood Lactate Concentrations," *JAMA*, vol. 272, no. 21, p. 1678, Dec. 1994, doi: 10.1001/jama.1994.03520210062033.
- [7] J. Toffaletti, "Elevations in blood lactate: Overview of use in critical care," *Scandinavian Journal of Clinical and Laboratory Investigation*, vol. 56, no. sup224, pp. 107–110, Jan. 1996, doi: 10.3109/00365519609088628.

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- [8] N. Baishya, M. Mamouei, K. Budidha, M. Qassem, P. Vadgama, and P. Kyriacou, "In-vitro spectrometric analysis of hyperlactatemia and lactic acidosis in buffer relating to sepsis," *Journal of Near Infrared Spectroscopy*, vol.29, no. 1, pp. 53–59, Feb. 2021, doi: 10.1177/0967033520968951.
- [9] D. Lafrance, "Measurement of lactate in whole human blood with nearinfrared transmission spectroscopy," *Talanta*, vol. 60, no. 4, pp. 635–641, Jul. 2003, doi: 10.1016/S0039-9140(03)00042-0.
- [10] K. Budidha, M. Mamouei, N. Baishya, M. Qassem, P. Vadgama, and P. A. Kyriacou, "Identification and Quantitative Determination of Lactate Using Optical Spectroscopy—Towards a Noninvasive Tool for Early Recognition of Sepsis," *Sensors*, vol. 20, no. 18, p. 5402, Sep. 2020, doi:10.3390/s20185402.
- [11] S. Chatterjee, K. Budidha, M. Qassem, and P. A. Kyriacou, "In-silico investigation towards the non-invasive optical detection of blood lactate," *Sci Rep*, vol. 11, no. 1, p. 14274, Dec. 2021, doi: 10.1038/s41598-021-92803-x.
- [12] V. Saptari and K. Youcef-Toumi, "Design of a mechanical-tunable filterspectrometer for noninvasive glucose measurement," *Appl. Opt.*, vol. 43, no.13, p. 2680, May 2004, doi: 10.1364/AO.43.002680.
- [13] R. A. Viscarra Rossel, "ParLeS: Software for chemometric analysis of spectroscopic data," *Chemometrics and Intelligent Laboratory Systems*, vol. 90, no. 1, pp. 72–83, Jan. 2008, doi: 10.1016/j.chemolab.2007.06.006.
- [14] P. Bhandare et al., "Multivariate Determination of Glucose in Whole Blood Using Partial Least-Squares and Artificial Neural Networks Based on Mid-Infrared Spectroscopy," *Appl. Spectrosc.*, AS, vol. 47, no. 8, pp.1214–1221, Aug. 1993, Accessed: Feb. 04, 2022. [Online]. Available: <https://opg.optica.org/as/abstract.cfm?uri=as-47-8-1214>